

Better Training for Safer Food BTSF

INSPECTION AND SAMPLING PROCEDURES

GIUSEPPE BOVO







OVERVIEW

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KHV SAMPLING
ISA SAMPLING
EUS SAMPLING



AIM OF THE INSPECTION

Inspection and sampling procedures of aquatic animal farms are different according to the purposes of the inspection:

- TO CONFIRM A SUSPICION
- TO DEMONSTRATE THE ABSENCE OF THE PATHOGEN IN ORDER TO ACHIEVE THE FREE STATUS



NATURE OF THE SUSPICION

Suspicion could be based on one or more of the following factors:

UNUSUAL MORTALITY

EVIDENCE OF CLINICAL SIGNS

WATER SOURCE BELONGING TO THE SAME RIVER CATCHMENT WHERE INFECTION WAS FIRST DETECTED

RECENT INTRODUCTION OF LIVE FISH FROM AN INFECTED SITE

RECENT INTRODUCTION OF FISH PRODUCTS FOR PROCESSING OR DISPOSAL

COMMON TRANSPORT SUCH AS LIVE AND DEAD FISH AND FISH FOOD DELIVERY

SHARED EQUIPMENTS WITH AN INFECTED SITE SUCH AS GRADERS, FISH BOXES, NEETS,





TEMPORARY RESTRICTIONS

Following a suspicion of a listed disease, waiting for the final lab results, the CA shall apply the following restrictions:

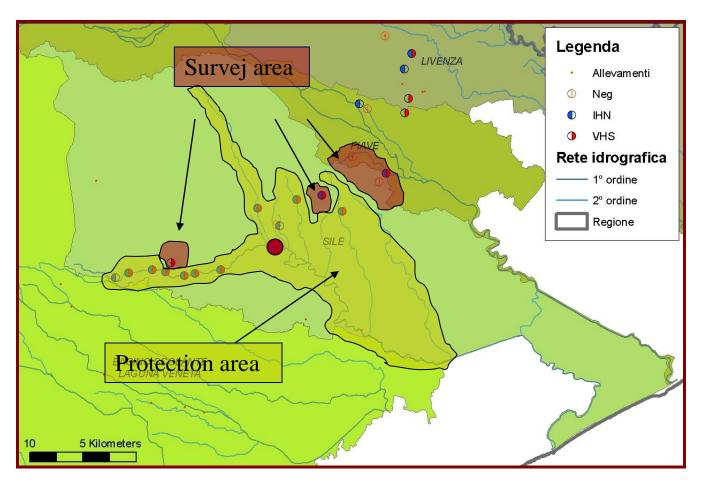
NO AQUACULTURE ANIMALS MAY LEAVE OR ENTER THE SITE WITHOUT PERMISSION AND CONTINUED SURVEILLANCE OF THE SITE IS REQUIRED

FURTHER SAMPLING MAY NEED TO BE UNDERTAKEN ON SITES IN CONTACT WITH THE COMPARTMENT UNDER SUSPICION IF AN EXOTIC DISEASE IS SUSPECTED

WHERE IT IS SHOWN THAT DISEASE MAY HAVE BEEN SPREAD TO OR FROM DIFFERENT SITES THESE SHOULD ALSO HAVE MOVEMENT CONTROLS APPLIED



CONTAINMENT AREA









EPIZOOTIC INVESTIGATION

The compartment under suspicion should be inspected for evidence of clinical disease in all fish holding facilities. An epizootic investigation shall be aimed at:

DETERMINE THE ORIGIN AND MEANS OF CONTAMINATION

INVESTIGATING WHETHER DURING THE RELEVANT PERIOD ANIMALS FROM THE SUSPECTED SITE HAS LEFT THE FARM

INVESTIGATING IF OTHER FARMS BECAME INFECTED





INSPECTION PROCEDURE -1

APPLY PERSONAL BIOSECURITY MEASURES TO AVOID CONTAMINATION

CONTROL THE EXISTENCE OF UPSTREAM/DOWNSTREAM COMPARTMENTS

CHECK RECORDS CONCERNING WATER QUALITY PARAMETERS (DISSOLVED OXIGEN, TEMPERATURE...) STOCKING DENSITY, RECENT STRESFULL EPISODES (GRADING, VACCINATION, NEW ARRIVAL, TREATMENTS), EXTERNAL VISITORS

CHECK THE PRESENCE OF ICHTHYOPHAGUS BIRDS

CHECK THE FARM REGISTER FOR:

SPECIES AND AGE OF FARMED FISH LIVE AND DEAD FISH MOVEMENTS RECORD OF DAILY MORTALITY RESULTS OF RISK-BASED SURVEILLANCE MORTALITY DURING TRANSPORT WATER EXCHANGE DURING TRANSPORT





INSPECT THE DEAD FISH STOCKING ROOM





INSPECTION PROCEDURE 2

INSPECT ALL THE UNITS FOR CLINICAL SIGNS, IN PARTICULAR INLET AND OULET WATER PLAICES:

LETHARGY
ABNORMAL SWIMMING BEHAVIOUR
SKIN DARKENING
SKIN HAEMORRHAGIES
SKIN RED DOTS OR EROSIONS
SWOLLEN ABDOMEN
EXOPTHALMUS
SUNKEN EYES
PERIORBITAL HAEMORRHAGIES
SKELETAL DEFORMITIES
RESPIRATORY PROBLEMS





INSPECTION PROCEDURE - 3

COLLECT SAMPLES (10 FISH) IN CASE OF CLINICAL SIGNS

COLLECT A LARGER SAMPLE (150 FISH) IN CASE OF ABSENCE OF SIGNS

CHECK AVAILABILITY OF DEAD FROZEN FISH

INTERVIEW THE RESPONSIBLE PERSON OF THE RISK-BASED SURVEILLANCE AND ALL THE PERSONS ATTENDING THE ANIMALS





SELECT THE RIGHT FISH





GENERAL SAMPLING PROCEDURE -1

IN CASE OF SUSPICION SAMPLES SHOULD BE TAKEN AMONG MORIBUND FISH SHOWING EVIDENT CLINICAL SIGNS

FRESH DEAD FISH ARE ALSO SUITABLE FOR DIAGNOSIS

IT IS RECOMMENDED TO PREPARE COLLECTIVE SAMPLES DIRECTLY IN THE FARM

USE ONLY STERILE TOOLS FOR COLLECTING TISSUES

WHOLE FISH ARE ALSO ACCEPTABLE (SENT ALIVE OR REFRIGERATED)

KEEP TISSUES, DEAD AND LIVING FISH AT <10° C









GENERAL SAMPLING PROCEDURES -2

IDENTIFICATION OF SAMPLE:

FARM CODE
SPECIES
AGE
TEMPERATURE
TANK IDENTIFICATION N°
CLINICAL DATA
ADDITIONAL NOTES















recommended n° fish in each collective sample

VHS/IHN: 10

KHV: 2

ISA:5

EUS: 1

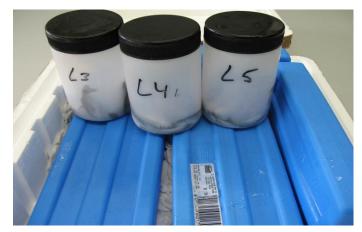




SHIPMENT OF SAMPLES









IHN & VHS SAMPLING -1 (Decision 2003/183/EC)

Sampling to confirm VHS/IHN under suspicion must be performed taking in consideration the following:

CLINICAL INSPECTION BETTER PERFORMED OCTOBER-JUNE OR AT TEMPERATURE < 14° C

INSPECT ALL PRODUCTION UNITS IN PARTICULAR NEAR TO WATER OUTLET

IF RAINBOW TROUT IS PRESENT COLLECT ONLY THIS SPECIES ON THE CONTRARY COLLECT DIFFERENT SPECIES IN %

SELECT ONLY FISH SHOWING CLEAR CLINICAL SIGNS (LETHARGY, EXOPTHALMUS, SKIN DARKENING, ERRATHIC SWIMMING)

IF FRESH DEAD FISH ARE AVAILABLE COLLECT THESE FISH PRIMARILY.



IHN & VHS SAMPLING -2

OVARIAN FLUID/TARGET TISSUES (0.5 GRAMS) FROM 10 FISH ARE COLLECTED AND SUSPENDED IN 4 ML OF TRANSPORT MEDIUM:

ANTERIOR KIDNEY
SPLEEN
HEART OR ENCEPHALON

WHOLE LIVING OR DEAD FISH ARE ALSO CONVENIENT PROVIDE TEMPERATURE IS MAINTAINED < 10° C

KEEP SAMPLES REFRIGERATED AT 4-10 ° C

THE LABORATORY MUST START INVESTIGATION < 48 HOURS POST SAMPLING.





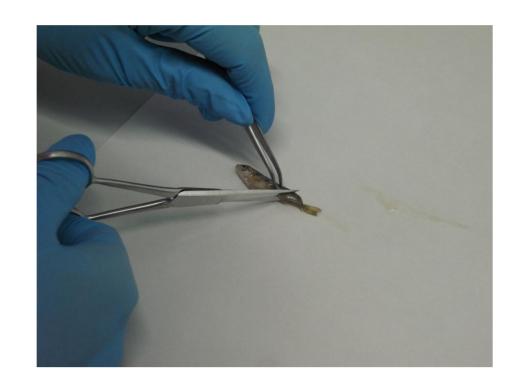
PREPARATION OF THE SAMPLES

FISH < 4 cm:

REMOVE THE BODY BEHIND THE ANUS

MINCE WITH STERILE SCISSORS

SUSPEND 1:10 IN TRANSPORT MEDIUM







PREPARATION OF THE SAMPLES -2

FISH 4-6 cm

COLLECT VISCERA INCLUDING KIDNEY

MINCE WITH STERILE SCISSORS

SUSPEND 1:10 IN TRANSPORT MEDIUM



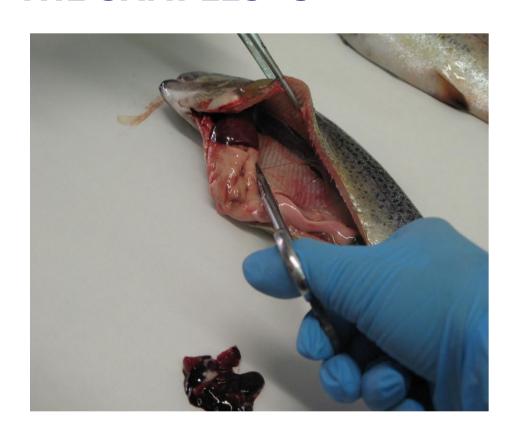


PREPARATION OF THE SAMPLES -3

FISH > 6 cm

COLLECT: SPLEEN
ANTERIOR KIDNEY
HEART/ENCEPHALON

SUSPEND 1:10 IN TRANSPORT MEDIUM

















FREEZING IN EXCEPTIONAL CASES

Where practical difficulties arise (eg. bad weather conditions, non-working days, laboratory problems) which make it impossible to inoculate cells within 48 hours after the collection of the tissue samples, it is acceptable:

TO FREEZE TISSUE SAMPLES IN TRANSPORT MEDIUM AT -20° OR BELOW

VIROLOGICAL EXAMINATION SHALL BE CARRY OUT < 14 DAYS

ONLY ONE THAWING/FREEZING CYCLE IS PERMITTED

RECORD MUST BE KEPT





ISA SAMPLING -1

(Dec. 2003/466/EC)

INSPECT ALL UNITS OF THE FARM (CAGES, TANKS, PONDS)

CHECK FOR FRESHLY DEAD, WEAK OR ABNORMALLY BEHAVING FISH

EXAMINE THOSE FISH FOR CLINICAL SIGNS AND POST MORTEM FINDINGS OF ISA

IF CLINICAL SIGNS ARE PRESENT COLLECT AT LEAST 10 FISH

IF CLINICAL SIGNS AND POSTMORTEM FINDINGS ARE NOT CONSISTENT WITH ISA SPECIFIC SAMPLING IS NOT NECESSARY BUT A DIFFERENTIAL DIAGNOSIS IS RECOMMENDED





ISA SAMPLING -2

(Dec. 2003/466/EC)

SAMPLES FOR HISTOLOGY

REMOVE SMALL PIECES (0.5X 0.5) OF LIVER, MID-KIDNEY, HEART AND SPLEEN AND FIX IN BUFFERED 10% FORMALIN (1 TISSUE: 20 FIXATIVE)

IMMUNOFLUORESCENCE

PREPARE IMPRINTS FROM KIDNEY TAKEN < 2 HOURS AFTER DEATH
ABSORBE BLOOD EXCESS ON ABSORBENT PAPER
MAKE LIGHT IMPRINTS ON POLY-I-LYSINE-COATED SLIDES
AIR DRY SLIDES AT ROOM TEMPERATURE
FIX < 72 HOURS OR STORE AT -20° C
SEND SLIDES TO THE LAB KEEPING DRY AND REFRIGERATED





ISA SAMPLING -2 (Dec. 2003/466/EC)

MOLECULAR TOOLS

REMOVE PIECES OF ANTERIOR KIDNEY OR MID-KIDNEY FROM EACH FISH AND TRANSFER INTO A MICROFUGE TUBE CONTAINING 1 ML OF RNAlater. KEEP REFRIGERATED/FROZEN

VIRUS ISOLATION

REMOVE PIECES OF LIVER, ANTERIOR KIDNEY, HEART AND SPLEEN (1 GRAM) AND TRANSFER INTO A TUBE WITH 9 ML TRANSPORT MEDIUM

PREPARE COLLECTIVE SAMPLES FROM < 5 FISH

KEEP SAMPLES REFRIGERATED UNTIL THEIR ARRIVAL AT THE LAB WHERE THEY SHALL BE PROCESSED < 72 HOURS

DUPLICATE SAMPLES SHALL BE TAKEN FOR CORROBORATIVE PURPOSES





KHV SAMPLING -1

SELECT 10 MORIBUND OR FRESHLY DEAD CARPS DISPLAYING TYPICAL CLINICAL SIGNS.

COLLECT TISSUES FROM

CLINICALLY INFECTED FISH (PCR, VIRUS ISOLATION)

- GILL, KIDNEY SPLEEN

SUBCLINICALLY INFECTED FISH (PCR)

- GILL, KIDNEY, SPLEEN, GUT, ENCEPHALON

POOLING OF SAMPLES SHOULD BE AVOIDED OR LIMITED TO A MAXIMUM OF TWO FISH PER COLLECTIVE SAMPLE





KHV SAMPLING -2

WHOLE FISH SHOULD BE SENT TO THE LABORATORY ALIVE OR KILLED AND PACKED SEPARATELY IN SEALED ASEPTIC CONTAINERS.

HOWEVER, IT IS HIGHLY PREFERABLE AND RECOMMENDED TO COLLECT ORGAN SAMPLES AT THE FISH PRODUCTION SITE.

WHOLE FISH OR SELECTED TISSUES SHOULD BE SENT TO THE LABORATORY IN REFRIGERATED CONTAINERS OR ON ICE

FREEZING OF FISH OR DISSECTED ORGANS SHOULD BE AVOIDED.





EUS SAMPLING

FISH SELECTION AND POOLING

SELECT 10 FISH WITH TYPICAL LESION IN <u>EARLY STAGES</u> OF INFECTION.

SEND SAMPLES TO THE LAB IN ICE-COOLED BOXES.

FISH COLLECTED FROM REMOTE AREAS SHOULD BE ANESTHETISED AND FIXED IN 10% BUFFERED FORMALIN FOR AT LEAST 1-2 DAYS.

DIAGNOSIS IS ACHIEVED USING THE HISTOLOGICAL TECHNIQUE AND OOMYCETE ISOLATION ON INDIVIDUAL FISH OR A GROUP OF A FEW FISH





THANK YOU FOR YOUR ATTENTION

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